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## THIOUREA ANALOGUES OF RESINIFERATOXIN AS LIGANDS FOR THE VANILLOID RECEPTOR

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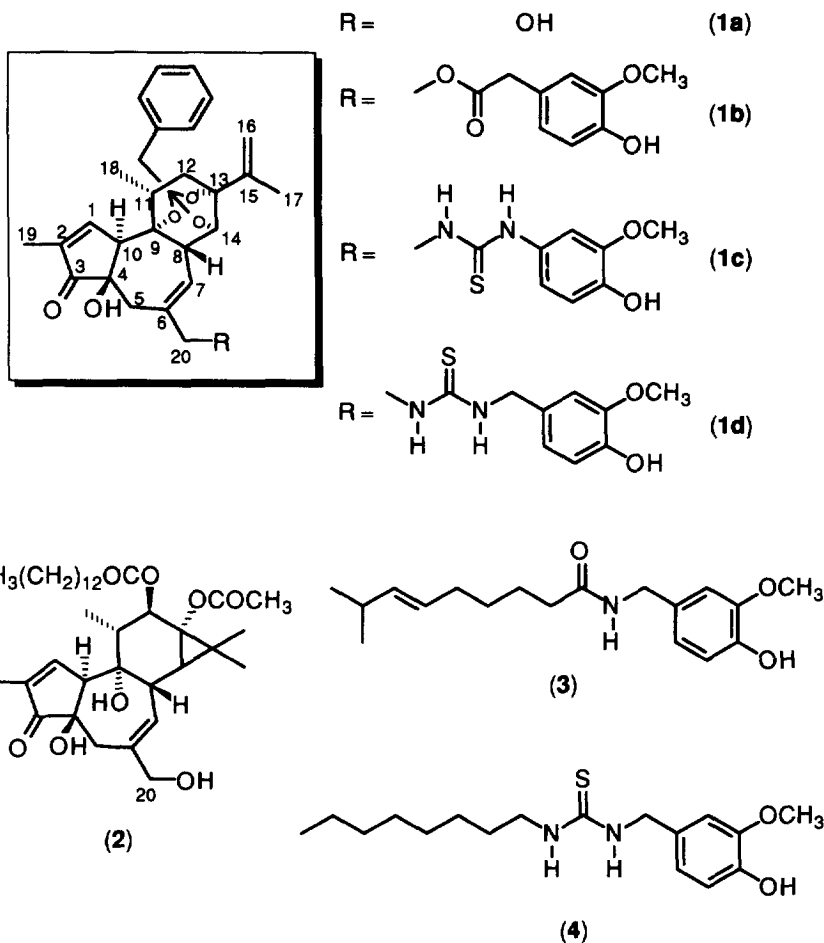
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**Abstract.** A thiourea bridge was tested as a linker to replace the OC(O)CH<sub>2</sub> ester group that connects resiniferonol-9,13,14-orthophenylacetate to the 3-methoxy-4-hydroxyphenyl moiety in resiniferatoxin (RTX). The resulting compounds showed lower affinity for the vanilloid receptor than did the parent RTX.

Resiniferatoxin (**1b**, RTX) is an extremely potent irritant tricyclic diterpene which is structurally related to other phorbol-related diterpenes, such as phorbol 12-myristate-13-acetate (**2**, PMA), except for its homovanillyl ester group at C-20.<sup>1,2</sup> In terms of irritancy, RTX is 100- to 1000-times more potent than PMA<sup>3</sup> but fails to induce other typical phorbol ester responses, and its affinity for protein kinase C is also much lower than that of the phorbol esters.<sup>4</sup> Instead, RTX appears to function pharmacologically as a "vanilloid", a group of compounds also characterized by their irritant properties that include the prototype capsaicin (**3**) and its analogues.<sup>5,6</sup> Capsaicin and its analogues comprise a class of potential non-narcotic, non-steroidal, analgesic-antiinflammatory agents which desensitize primary sensory afferent neurons. One of the functions of these neurons is to transmit nerve impulses to the central nervous system which are perceived as pain.<sup>7</sup> RTX and capsaicin share a common vanilloid pharmacophore which is essential for the extreme potency of RTX. Indeed, resiniferonol-9,13,14-orthophenylacetate (**1a**, ROPA), the 20-deacylated parent structure, fails to bind to the RTX receptor.<sup>8,9</sup> Both RTX and capsaicin bind to the same receptor, but RTX displays 1000- to 10000-fold greater potency than capsaicin, thus functioning as an ultrapotent capsaicin analogue.<sup>5</sup> Structure-activity studies on capsaicin analogues<sup>10-13</sup> have established that the mid-region, comprising the amide bond that connects the aromatic 3-methoxy-4-hydroxyphenyl moiety to the aliphatic side chain, can tolerate an ester function and even the reversal of the amide bond without seriously compromising activity.<sup>11</sup> From this study, a thiourea bridge, as in *N*-(4-hydroxy-3-methoxybenzyl)-*N'*-octylthiourea (**4**), provided the most potent compound of the series.<sup>11</sup> Armed with this information, we wished to modify the RTX structure with a similar thiourea bridge connecting the aromatic region and the diterpene moiety to give compounds **1c** and **1d**. These compounds were of further interest in that they would be expected to be metabolized differently from RTX. Hydrolysis of the ester in RTX yields ROPA, a relatively weak protein kinase C agonist, whereas hydrolysis of the ureas would not only be more difficult, but it would generate the amine analogue of ROPA (**12**), a compound that we have shown previously to have very poor affinity for protein kinase C.<sup>14</sup> Since structure-activity studies on capsaicin have also indicated the importance of the length of the mid-region, this issue was considered by attaching the thiourea bridge either directly to the aromatic ring as in **1c**, or

through a methylene spacer as in **1d**.

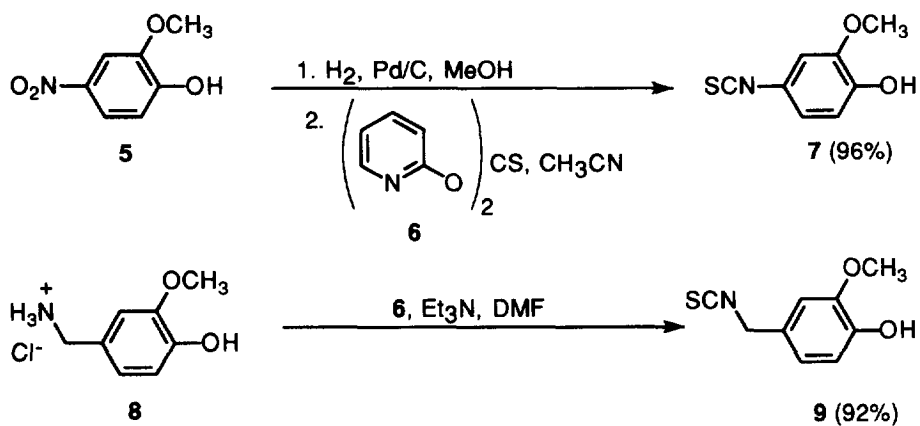
The requisite isothiocyanates were prepared according to Scheme 1. Reduction of commercially available 4-nitroguaiacol (**5**) provided the intermediate amine which upon treatment with di-2-pyridylthionocarbonate<sup>15</sup> afforded isothiocyanate **7**. For the higher homologue, commercially available 4-hydroxy-3-methoxybenzyl amine hydrochloride was reacted with the same reagent in the presence of



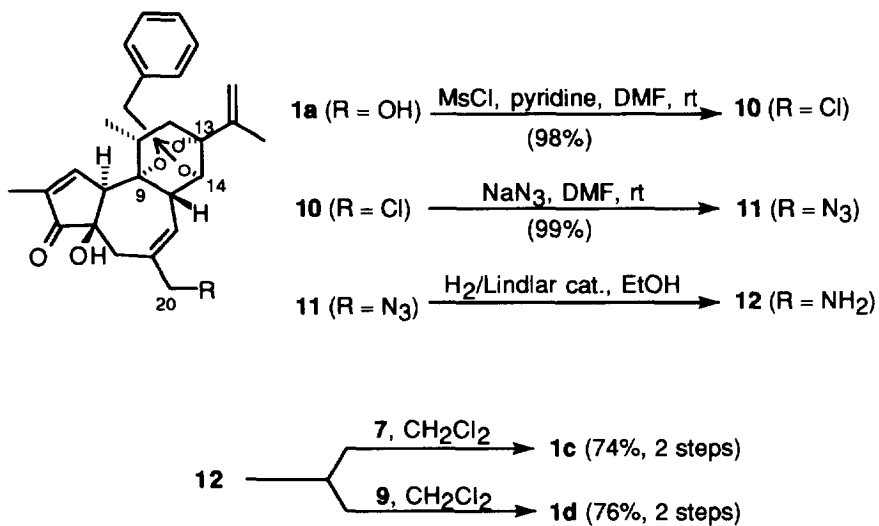
triethylamine to give isothiocyanate **9**. By using known phorbol chemistry,<sup>16</sup> ROPA (**1a**) was converted to the corresponding chloride **10** after treatment with methanesulfonyl chloride (Scheme 2). Displacement with  $\text{NaN}_3$  gave the corresponding azide **11**, which was then reduced in the presence of Lindlar catalyst<sup>17</sup> to the amine **12**. This amine was used directly in condensation reactions with isothiocyanates **7** and **9** to give the final products (Scheme 2).<sup>18,19</sup>

The capsaicin-like activities of these RTX analogues were measured by *in vitro* binding experiments

Scheme 1.



Scheme 2.



in which specifically bound [ $^3\text{H}$ ]RTX was displaced by the non-radioactive competing ligands. The specific details of this method have been published.<sup>14</sup> Activity is expressed in terms of a  $K_i$  value (mean  $\pm$  SEM, 3 experiments) which represents the concentration of the non-radioactive ligand which displaces half of the specifically bound labelled RTX, corrected for RTX concentration and RTX affinity at the receptor. The values obtained for **1c** ( $K_i = 66.1 \pm 0.8$  nM) and for **1d** ( $K_i = 149 \pm 11$  nM) indicate that these compounds are, respectively,  $3 \times 10^3$  and  $6.5 \times 10^3$  less potent than RTX ( $K_i = 23.1 \pm 3.1$  pM). We can conclude that despite having a common "vanilloid" pharmacophore, a simple juxtaposition of an "optimized" capsaicin segment on the resiniferonol-9,13,14-orthophenylacetate moiety does not produce a better ligand. Changes in the homovanillyl portion of RTX, as the ones described here, presumably disrupt the alignment between the diterpene moiety and the homovanillyl ester group at C-20 that contributes to the compound's ultra-high potency.

## References and Notes

- Adolph, W.; Sorg, B.; Hergenbahn, M.; Hecker, E. *J. Nat. Prod.* **1982**, *45*, 347.
- Schmidt, R. J.; Evans, F. J. *Inflammation* **1979**, *3*, 273 (1979).
- Hecker, E. *Carcinogenesis* **1978**, *2*, 11.
- Driedger, P.; Blumberg, P. M. *Cancer Res.* **1980**, *40*, 1400.
- Szallasi, A.; Blumberg, P. M. *Neuroscience* **1989**, *30*, 515.
- de Vries, D.; Blumberg, P. M. *Life Sci.* **1989**, *44*, 711.
- Szallasi, A.; Blumberg, P. M. *Adv. Pharmacol.* **1993**, *24*, 123.
- Szallasi, A.; Blumberg, P. M. *Brain Res.* **1990**, *524*, 106.
- Szallasi, A.; Sharkey, N.; Blumberg, P. M. *Phytotherapy Res.* **1989**, *3*, 253.
- Walpole, C. S. J.; Wrigglesworth, R.; Bevan, S.; Campbell, E. A.; Dray, A.; James, I. F.; Perkins, M. N.; Reid, D. J.; Winter, J. *J. Med. Chem.* **1993**, *36*, 2362.
- Walpole, C. S. J.; Wrigglesworth, R.; Bevan, S.; Campbell, E. A.; Dray, A.; James, I. F.; Masdin, K. J.; Perkins, M. N.; Winter, J. *J. Med. Chem.* **1993**, *36*, 2373.
- Walpole, C. S. J.; Wrigglesworth, R.; Bevan, S.; Campbell, E. A.; Dray, A.; James, I. F.; Masdin, K. J.; Perkins, M. N.; Winter, J. *J. Med. Chem.* **1993**, *36*, 2381.
- Janusz, J. M.; Buckwalter, B. L.; Young, P. A.; LaHann, T. R.; Farmer, R. W.; Kasting, G. B.; Loomans, M. E.; Kerckaert, G. A.; Maddin, C. S.; Berman, E. F.; Bohne, R. L.; Cupps, T. L.; Milstein, J. R. *J. Med. Chem.* **1993**, *36*, 2595.
- Acs, G. Lee, J.; Marquez, V. E.; Wang, S.; Milne, G. W. A.; Du, L.; Lewin, N. E.; Blumberg, P. M. *J. Neurochem.* in press.
- Kim, S.; Yi, K. Y. *Tetrahedron Lett.* **1985**, *26*, 1681.
- Thielmann, H. W.; Hecker, E. *Lieb. Ann. Chem.* **1969**, *728*, 158.
- Corey, E. J.; Nicolau, K. C.; Balanson, R. D.; Machida, Y. *Synthesis* **1975**, 590.
- Compound **1c**: white solid, mp 148–50 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.66 (br s, 1 H, S=CNH-Ar), 7.42 (s, 1 H, H-1), 7.10–7.40 (m, 5 H, Ph), 6.70–7.00 (m, 3 H, Ar), 5.60–6.00 (m, 2 H, OH, H-7), 4.68 (br s, 2 H, H-16), 4.05–4.50 (m, 3 H, H-14, H-20), 3.84 (s, 3 H,  $\text{OCH}_3$ ), 3.00–3.23 (m, 4 H,  $\text{PhCH}_2$ , H-8, H-10), 2.00–2.60 (m, 5 H, H-5, H-11, H-12), 1.78 (br s, 3 H, H-19), 1.50 (br s, 3 H, H-17), 0.92 (d, 3 H, H-18); FAB MS ( $m/z$ , relative intensity) 645.6 ( $\text{MH}^+$ , 23).
- Compound **1d**: white solid, mp 144–46 °C;  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.45 (s, 1 H, H-1), 7.15–7.38 (m, 5 H, Ph), 6.90–7.05 (m, 3 H, Ar), 5.98 (br s, 1 H, OH), 5.90 (br s, 1 H, NH), 5.42 (s, 1 H, H-7), 4.26–4.90 (m, 4 H, H-16,  $\text{NHCH}_2\text{Ar}$ ), 3.70–4.10 (m, 6 H,  $\text{OCH}_3$ , H-20, H-14), 2.50–3.20 (m, 6 H,  $\text{PhCH}_2$ , H-8, H-10, H-5), 1.96–2.18 (m, 3 H, H-11, H-12), 1.78 (br s, 3 H, H-19), 1.56 (br s, 3 H, H-17), 0.97 (d, 3 H, H-18); FAB MS ( $m/z$ , relative intensity) 659 ( $\text{MH}^+$ , 24).